Transactions of the American Fisheries Society

parameters in bioenergetics models for juvenile Chinook salmon

22 **ABSTRACT**

Fsnawysma produced estimates of maximum consumption (C_{max}) at an measured in published laboratory feeding trails. In this paper w
unpublished data from laboratory feeding trails. In this paper w
unpublished data from l 23 Researchers have cautioned against the borrowing of consumption and growth parameters from 24 other species and life stages in bioenergetics growth model projections. In particular, 25 temperature dependent consumption in the Wisconsin bioenergetics model for Chinook salmon 26 *Oncorhynchus tshawytsha* produced estimates of maximum consumption (C_{max}) at lower 27 temperatures than measured in published laboratory feeding trails. In this paper we used 28 published and unpublished data from laboratory feeding trials for three stocks of subyearling 29 Chinook salmon: the Snake/Columbia River, the Nechako/Fraser River, and the Big Qualicum 30 River to estimate and adjust parameters for temperature dependence in *C*max used in the 31 Wisconsin bioenergetics model. Our data included growth measures from fish ranging from 1.5 32 to 7.2 g, and at temperatures that ranged from 14 to 26°C. We estimated parameters for 33 temperature dependence in *C*max based on relative differences in food consumption and then used 34 bootstrapping techniques to estimate the error about the parameters. We found that the current 35 parameter values miss-estimated the observed data at temperatures between 17 and 25 \degree C, 36 confirming that the C_{max} should be shifted by about 4° C relative to the current implementation 37 under the bioenergetics model. We conclude the adjusted parameters for *C*max should produce 38 more accurate predictions from the bioenergetics model for subyearling Chinook salmon.

40 **INTRODUCTION**

For Properties and Anderson 2011), better understand nutrition

For Produkte aquaculture systems (Jobling 1994; Dumas et al. 2009), a

Fehaviors in the wild (Armstrong and Schindler 2011). These models

using empirical dat 41 Bioenergetics models have become increasingly important tools in fisheries biology to examine 42 the effects of climate change (Mruscia et al. 2009; Pőrtner and Peck 2010; Gale et al. 2011; 43 Hasler et al. 2012), predict the consequences of habitat alterations and restorations (Boughton et 44 al. 2007; Wehrly et al. 2007; Beer and Anderson 2011), better understand nutritional physiology 45 and toxicology, evaluate aquaculture systems (Jobling 1994; Dumas et al. 2009), and compare 46 consumptive behaviors in the wild (Armstrong and Schindler 2011). These models have been 47 parameterized using empirical data from individuals or groups of fish across a wide range of 48 species, locations, and life stages (Kitchell et al. 1977; Beauchamp et al. 1989; Stewart and 49 Ibarra 1991; Rand et al. 1993; Railsback and Rose 1999; Tyler and Bolduc 2008). In 50 constructing bioenergetics models, parameters for the population of interest are often borrowed 51 from other species, locations, or life stages due to lack of data about the target population. 52 However, such borrowing of parameters may introduce bias into growth or consumption 53 predictions from bioenergetics models (Ney 1993; Chipps and Wahl 2008; Tyler and Bolduc 54 2008; Dumas et al. 2009; Van Poorten and Walters 2010). Researchers have also cautioned that 55 parameters derived from older or larger fish may not accurately fit juvenile stages (Hanson et al. 56 1997; Tyler and Bolduck 2008). Juvenile fish often have different temperature-dependent 57 consumption and growth relationships compared to adults of the same species (Post 1990; King 58 et al. 1999; Schoenbeck et al. 2008; Ohlberger et al. 2012), and thus borrowing parameter values 59 from other species or life stages may lead to systematic departures in bioenergetics estimates 60 (Bajer et al. 2004; Trudel et al. 2005). 61 In bioenergetics models, the purpose of the function for temperature dependence in

62 maximum consumption (*C*max; Thornton and Lessem 1978; Hanson et al. 1997) is to scale the

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134 Following the notation of Hanson et al. (1997) K_A and K_B can be expressed as:

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K_A = (CKI \cdot LI) / (1 + CKI \cdot (LI-1))
$$

$$
136 \t\t Ll = e^{(GI \cdot (T-CQ))}
$$

137 *G1* = (1/(*CTO*-*CQ*))·ln((0.98·(1-*CK1*))/(*CK1*·0.02))

138 $K_B = (CK4 \cdot L2)/(1+CK4 \cdot (L2-1))$

- 139 $L2 = e^{(G2 \cdot (CTL T))}$
- 140 *G2* = (1/(*CTL*-*CTM*))·ln((0.98·(1-*CK4*))/(*CK4*·0.02))
- 141

 $E_2 = e^{\epsilon}$
 $G_2 = (1/(CTL - CTM)) \cdot \ln((0.98 \cdot (1 - CK4))/(CK4 \cdot 0.02))$

mships *T* is the water temperature. For the increasing portion of the

temperature at which temperature dependence is a small fraction (is

sumption rate; and *CTO* 142 In these relationships *T* is the water temperature. For the increasing portion of the curve; *CQ* is 143 the lower water temperature at which temperature dependence is a small fraction (*CK1*) of the 144 maximum consumption rate; and *CTO* is the water temperature that is 0.98 of the maximum 145 consumption rate. Similarly, for the decreasing portion of the curve, *CTM* is defined as the water 146 temperature at which dependence is still 0.98 of the maximum, and *CTL* is the temperature at 147 which dependence is some reduced fraction (*CK*4) of the maximum consumption rate. Please 148 see Thornton and Lessem (1978) for greater detail on the general construct of this model for 149 biological rates.

150 We used the food consumption data provided by Brett et al. (1982) and collected during 151 our laboratory studies to estimate the six consumption-dependent parameters used by Thornton 152 and Lessem (1978) (Tables 1 and 2). Consumption data for the *j*th tank or treatment group of 153 tanks within an experiment were transformed to a scale ranging from 0 to 1as $\acute{C} = C_{\text{tot}i}/\sqrt{2}$ 154 max(C_{totij}) where C_{totij} is the total amount food eaten during the *i*th study by the *j*th tank or 155 treatment group over a growth period (Legendre and Legendre 1998). Thus when $\acute{C} = 1$ then 156 $C_{\text{totii}} = \max(C_{\text{totii}})$. Consumption data collected by Brett et al. (1982) were collected under fairly 157 constant temperatures. To maintain comparability among studies and fulfill the assumption of a

158 fixed *x*-axis, we used only the consumption data when temperatures were constant and within 159 1°C of the experimental target temperature (day 34 to 80 in 2003 and day 21 to 42 in 2004). 160 These *Ć* values and the associated mean tank temperatures were used to estimate the Thornton 161 and Lessem (1978) parameters. To provide a measure of uncertainty about the parameter values 162 we bootstrapped the residuals about the fitted line, and used the percentile method to obtain the 163 95% confidence limits (CL) about the parameter values (Efron and Tibsharini 1993).

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165 **RESULTS**

For Personals about the fitted line, and used the percentile method

For Personals (CL) about the parameter values (Efron and Tibsharini 199;

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For P 166 The amount of food consumed varied among tanks and studies, largely due to differences in the 167 length of the growth periods, fish sizes, and the numbers of fish (Table 1). For example, the total 168 amount of food eaten (g) over the 28-d Brett et al. (1982) study ranged from 32.1g to 116 g for 169 the Qualicum River fish, and 58.1g to 94.9 g for Nechako River fish. The amount of food 170 consumed by Snake River stocks ranged from 511 g to 994 g over our 21-d feeding trails, and 171 1175.0 g to 4007g over our 46-d feeding trails. The temperature at which the maximum amount 172 of food was eaten occurred at mean daily temperatures of about 20.9 °C for Qualicum River fish 173 and 21.0 °C for Nechako River fish in the Brett et al. (1982) studies. During our studies, 174 consumption (g/g/d) was maximized at mean temperatures of 16^oC to 20^oC, though we only 175 measured consumption at three temperatures. 176 Parameter estimates from fitting $f(T)$ to the consumption data supported a shift in 177 consumption towards higher temperatures than currently specified in the bioenergetics model for 178 Chinook salmon (Table 2 and Figure 1). The unadjusted model specified C_{max} at 16.7 °C, 179 whereas the adjusted parameter values estimated C_{max} at 20.9 °C (lower 95% CL =18.2, Upper 180 95% CL 21.8 =), a 4.2 -°C difference between models. This difference between the unadjusted

181 and adjusted model was consistent over the temperature range, although we had little data on 182 which to inform $f(T)$ at low temperatures, and so there was large uncertainty about the parameter 183 values at low end of the temperature range. Nonetheless, the estimated curve for temperature 184 dependence in maximum consumption more closely followed the observed data than did *f*(*T*) 185 from the unadjusted model. For example, the unadjusted model resulted in an $r^2 = 0.24$ and residuals that were heterogeneous over the predicted range, whereas the adjusted model had an *r* 2 186 $187 = 0.77$ and homogeneous residuals over the predicted range. These findings clearly support a 188 better fit to the data for the adjusted model and a shift towards C_{max} at higher temperatures for 189 subyearlings than currently implemented under the Wisconsin bioenergetics model for Chinook 190 salmon.

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192 **DISCUSSION**

Islea model. For example, the unadjusted model resulted in an $V =$

Fore heterogeneous over the predicted range, whereas the adjusted nogeneous residuals over the predicted range. These findings clearly

data for the adjus 193 Our re-evaluation of the Thornton and Lessem (1978) parameters should improve consumption 194 and growth estimates when using the Wisconsin based bioenergetics model in application to 195 juvenile Chinook salmon. The unadjusted parameters developed for Chinook salmon by Stewart 196 and Ibarra (1991) used lower optimal temperatures in the thermal multiplier equation for *C*max 197 from Thornton and Lessem (1978). We found that adjustments in the thermal multiplier 198 parameters better accounted for the effect of temperature on C_{max} across the range of rearing 199 temperatures for which we had data. However, we acknowledge that all fish in these 200 experiments were fed high energy, easily digestible diets, which potentially influenced our 201 results. Nonetheless, if our findings about juvenile feeding behavior at specific temperatures in 202 the laboratory are at all comparable to performance in the natural environment, use of our 203 adjusted model parameters should produce better estimates of consumption and growth for

204 juvenile fall Chinook salmon (Koehler et al. 2006; Armstrong and Schindler 2011).

The wisconsin model were developed without consumption data from

finfluencing the variable performance of this model (Mandenjian et a

For the unadjusted model parameter values for the temperature-dependent

For Parachamp 205 With the paucity of carefully collected data, and the difficulty in evaluating and obtaining 206 these key bioenergetics model parameters (Chips and Wahl 2008), we believe model predictions 207 can be improved for subyearling Chinook salmon with adjustments. The unadjusted model 208 parameters of the Wisconsin model were developed without consumption data from Chinook 209 salmon, likely influencing the variable performance of this model (Mandenjian et al. 2004; Chips 210 and Wahl 2008). The unadjusted model parameter values for the temperature-dependence in 211 *C*max were calibrated by Beauchamp et al. (1989) for sockeye salmon, *O. nerka.* Stewart and 212 Ibarra (1991) later compared the Beauchamp et al. (1989) calibration using food consumption by 213 coho salmon, *O. kisutch* (Edsall et al. 1974, 1999; Stewart 1980; Stewart et al. 1983), but not 214 Chinook salmon. So using existing data to re-parameterize the Thornton and Lessem (1978) 215 model for *C*max, provided a relatively inexpensive opportunity to evaluate and improve a sub-216 model to the Wisconsin bioenergetics model over a wide range of conditions know to be 217 important to fish consumption, and in turn growth estimates. 218 The early life stage and smaller fish sizes likely contributed to our results. Madenjian et

219 al. (2004) found good agreement between observed and predicted values for the unadjusted 220 bioenergetics model for Chinook salmon, suggesting an appropriate function for *C*max. However, 221 the authors conducted their laboratory evaluation using much larger Chinook salmon ($>$ 400 g) 222 that were fed natural (i.e, alewife, *Alosa pseudoharengus*) rather than pelleted feed, and fish were 223 reared at a cooler and narrower temperature range (10.7–13 $^{\circ}$ C) than evaluated by this study. 224 Sauter et al. (2001) experimentally showed that the thermal preference of subyearling fall 225 Chinook salmon (from the lower Columbia River) decreased from about 18 to 11°C with the 226 progression of smoltification, indicating that optimal temperatures for consumption (and growth)

249 parameter for consumption at the lower end of thermal range (< 10 °C). Given the *post-hoc*

248 tests designed to calibrate and corroborate the model parameters, especially those for *CQ* – the

273 **ACKNOWLEDGEMENTS**

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415 **Figure Captions**

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- 417 Figure 1. Adjusted (upper panel) and unadjusted (lower panel) temperature dependent
- 418 consumption curves using the Thornton and Lessem (1978) equation and coefficients provided in
- 419 Table 1. Individual plots of the proportion of maximum food consumption by mean water
- 420 temperatures are provided for data from laboratory trials of Brett et al (1982) for Qualicum River
- 421 and Nechako River stocks, and Snake River stocks reared in our laboratory. Dashed lines in the
- 422 upper panel represent the bootstrapped 95% confidence bounds about the adjusted consumption

423 curve.

424

472 Table 1. Mean tank temperatures, initial weights, and the total food consumed by subyearling

473 Chinook salmon by river origin, length of feeding trial at constant temperature, and data source.

474 Ranged food values were calculated from dry weights during the Brett et al. (1982) studies and

475 calculated from wet weights during U of Idaho studies.

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